

Stable amino acid based bicarbonate solution for periton al dialysis and h modialysis

Patent Number: EP1166787
Publication date: 2002-01-02
Inventor(s): YATZIDIS HIPPOCRATES (GR)
Applicant(s):: YATZIDIS HIPPOCRATES (GR)
Requested Patent: EP1166787
Application Number: EP20010600017 20010614
Priority Number(s): GR20000100214 20000628
IPC Classification: A61K33/10 ; A61K31/194 ; A61K31/198 ; A61K31/405
EC Classification:
Equivalents: GR1003567

Abstract

The invention relates to two single and stable solutions for peritoneal dialysis (PD) and hemodialysis (HD). The solutions contain the needed electrolytes, the bicarbonate and at very small concentration (1.5mM/L) disodium hydrogen citrate. Besides, the PD solution contains the following ten indispensable amino acids, L-histidine, L-leucine, L-lysine, L-methionine, L-phenylalanine, L-threonine, L-tryptophane, L-valine, L-arginine and L-glutamine, while the HD solution contains the two dispensable amino acids, L-aspartic acid and L-glutamic acid.

Data supplied from the esp@cenet database - I2

Description

Background of The Invention

[0001] Peritoneal dialysis (PD) and hemodialysis (HD) are commonly used methods in the treatment of patient with end-stage renal disease. The uremic patient's blood, is brought into contact with dialysis solutions across either the peritoneal or an artificial membrane and is purified from the waste products based on the principles of dialysis.

[0002] Such solutions should have an electrolyte formula resembling as closely as possible that of normal plasma and contain, for example, sodium, potassium, chloride, calcium and magnesium ions. In PD solutions glucose is used as an osmotic agent while lactate, acetate and bicarbonate ions as buffering agents. All currently used PD solutions still have many drawbacks with serious consequences, arising mainly from lactate, the low pH (5.2) and the high glucose concentrations and in HD solution from acetate.

[0003] At present three PD solutions containing glucose concentrations 15,25 and 42.5 g/L are used. Because of their high glucose concentrations, patients on continuous ambulatory peritoneal dialysis (CAPD) absorb daily about 200 g of glucose. This superabundant amount of glucose leads to hyperglycemia, hyperinsulinemia, hyperlipidemia and obesity, which are important risk factors for accelerating atheromatosis and arteriosclerosis. In addition, as these peritoneal solutions must be sterilized by autoclaving at a pH around 5.2 in order to avoid the caramelisation, toxic substances are generated such as glucose byproducts (5 hydroxyl methyl furfural, furandic carboxylic acid, acetyl acrylic acid), advanced glycosylation end products (AGEs), and reactive oxygen species (ROS) which damage peritoneum and its function.

[0004] On the other hand, acetate in HD solution (dialysate) is unable to control metabolic acidosis and furthermore has the potential of causing bicarbonate

[0005] On the other hand, acetate in HD solution (dialysate) is unable to control metabolic acidosis and furthermore has the potential of causing bicarbonate deficit, accumulation of acetate and its metabolites, as well as for consequent acid-base and hemodynamic disturbances. Bicarbonate has many advantages as it is the major natural buffer, with the better interdialytic hemodynamic stability and tolerance, can restore effectively acidosis with beneficial effects on protein metabolism, nutrition, the skeleton and the overall well-being of patients. However, there is a serious problem, as bicarbonate could react with calcium and magnesium and form insoluble neutral calcium and magnesium carbonate salts.

[0006] In order to overcome this drawback one approach uses two different solutions, one alkaline, containing bicarbonate, and the other acidic, containing calcium and magnesium, which are mixed on-line during dialytic therapy. A similar technique uses dry bicarbonate in a cartridge belt. It has been proposed, but not strongly supported, that in the on-line mixed solution, which ordinary has a pH of 6.8-6.9, insoluble neutral calcium and magnesium carbonate are formed late during infusion. This is correct, however, only when the solution is stored in the laboratory, including into air tight bottles, but not during treatment. In fact, studying this on-line solution at a dialyzer exit, after its contact with patient's blood through the artificial membrane, we noted that its pH reached 7.15-7.20, and it is deteriorated rapidly. More problematic is the use of on-line technique in CAPD. In fact, this is very complicated, tedious and risky as it necessitates 4 exchanges of sterile peritoneal solutions each day which are performed by the patient.

[0007] The on-line technique produces a non authentic bicarbonate solution as it becomes unstable during infusion and may decrease bicarbonate and favor the accumulation in the patient's plasma of carbonic acid, carbon dioxide, soluble hydrogen calcium and magnesium carbonates and finally insoluble neutral calcium and magnesium carbonate, the deposition of which, become evident on the wall of the connecting tubes (1-6).

[0008] From many years we have worked on the preparation of single bicarbonate solutions for PD and HD. Such improved single bicarbonate solutions have been produced by rendering them true and efficient buffers with pH 7.35 by adding a quantity of glycylglycine. At this constant pH the formation and precipitation of insoluble neutral calcium and magnesium carbonate are suspended. Bicarbonate solutions were patented internationally by the author and Pierre Fabre Medicament Society (Paris, France). Numerous experimental and clinical trials have demonstrated their superiority, compared to similar standard lactate and on-line produced bicarbonate solutions (7-11). The high cost of glycylglycine and the need for sterilization by filtration, not accepted in many countries have made their production unacceptable.

[0009] To overcome this hindrance we have replaced successfully glycylglycine by the L-aspartic and

L-glutamic acids at very low concentrations and prepared single and stable bicarbonate solutions, which are already patented in Greece and European Union (12).

[0010] We describe here the production of a single amino acid-based bicarbonate PD solution and a single bicarbonate HD solution by using simple, competent and safe technical approach.

Description of the solutions

[0011] The particular features of single bicarbonate solutions for both PD and HD are the inclusion of disodium hydrogen citrate into their formulation, plus the inclusion of amino acids into the PD solution.

[0012] Studies in our laboratories proved that the inclusion of disodium hydrogen citrate at a very low concentration (1.5 mM/L) into the PD and HD solutions of a pH 7.25 -7.45, completely abolishes the well known reaction between bicarbonate and calcium-magnesium. As citrate disposes a potent chelating capability on divalent cations, calcium and magnesium form with citrate water soluble complexes. Consequently, after the entry of these complexes in the circulation, citrate is rapidly metabolized to bicarbonate in the liver and the released calcium and magnesium regain their pure forms. We used disodium hydrogen citrate because may control the indicated pH and also because it is more stable than trisodium citrate. Amino acids added in PD solution instead of glucose.

Single amino acid- based bicarbonate PD solution

[0013] The exact formulation of single amino acid-based bicarbonate PD solution is listed in Table 1.

[0014] In 1968 Dudrick et al (13) introduced amino acids in parenteral i.v. nutrition and the same year Gjessing (14) added amino acids to the peritoneal solution, in an attempt to compensate for losses occurring during dialysis. In 1979 Oreopoulos et al used, for the first time, amino acids as osmotic agent in PD solution, at a concentration of 1 and 2%, instead of 1.5 and 4.25% glucose (15). Since then, hundreds of relative articles have been published.

Id=Table 1. Columns=5

Title: A single and stable improved bicarbonate amino acid solution for peritoneal dialysis

Head Col 1 to 2: Amino acids

Head Col 3: mw

Head Col 4: mM/L

Head Col 5: g/L

L-Histidine C₆H₉N₃O₂ 155.18 6 0.931

L-Leucine C₆H₁₃NO₂ 131.17 18 2.098

L-Lysine.HCl C₆H₁₄N₂O₂.HCl 182.60 10 1.826

L-Methionine C₅H₁₁NO₂S 149.21 8 1.193

L-Phenylalanine C₉H₁₁NO₂ 165.19 3 0.495

L-Threonine C₄H₉NO₃ 119.12 13 1.548

L-Tryptophan C₁₁H₁₂N₂O₂ 204.22 2 0.408

L-Valine C₅H₁₁NO₂ 117.15 18 2.108

L-arginine C₆H₁₄N₄O₂ 174.20 3 0.523

L-Glutamine C₅H₁₀N₂O₃ 146.15 15 2.192

Number of amino acids : 10 1544.17 94 13.322

Average mw : 154.42

Head Col 1 to 2: Electrolytes

Head Col 3:

Head Col 4:

Head Col 5: g/L

Sodium chloride NaCl 58.44 5.569

Sodium bicarbonate NaHCO₃ 84.01 2.940

di-sodium citrate C₆H₆Na₂O₇ 11/2 H₂O 236.09 0.354

Calcium chloride CaCl₂.2H₂O 146.98 0.221

Magnesium chloride MgCl₂.6H₂O 203.21 0.102

Na 135+ Cl 101.5+ HCO₃ 35+

Ca 1.5 + Ca 1.5+Mg 0.5 mM/L : 278

pH : 7.25-7.45 Collective osmolarity :389 mOsm

[0015] Amino acids are the structural units from which proteins are synthesized. They are unevenly distributed in the media of the body and are chemically reactive. Because amino acids possess a basic amino group they have the power to combine with either alkali or acid, acting as buffers. Amino acids can be classified, according to their chemical structures and reaction into 3 groups, in basic amino acids in which the ratio between basic amino group and acid carboxyl group is 2:1, in neutral amino acids in which the ratio is 1:1 and in acidic amino acids in which this ratio is 1:2. Dietary amino acids are a good source of energy in some organs. Thus almost 50% of oxygen consumed by the liver is used for amino acid oxidation, and in small intestine is almost 30%. In the muscle and kidney, however these figures are 7.5% and 12.5%, respectively. Not all amino acids or proteins are equal nutritionally. Negative nitrogen balance may occasionally have to be corrected by hyperalimentation or total parenteral nutrition (TPN) amino acids, plus a source of calories in the form of fat and carbohydrate and also contain all other nutritional factors required for life such as vitamins, and minerals. Early was discovered by Osborne and Mendel (16), that the elimination of certain amino acids from the diets prevented survival or growth, while the omission of others had no deleterious effect upon the body function. This had led to the classification of amino acids as essential or nonessential (17,18). These terms are less than precise. Today, it is preferred to use the terms "indispensable" and "dispensable" amino acids. In 1987, Laidlaw and Kopple (19) in a study in healthy adult human identified nine amino acids (isoleucine, leucine, lysine, methionine, phenylalanine, threonine, tryptophan, valine and histidine) which were shown, by classical nitrogen balance studies, to be indispensable, while the others were dispensable.

[0016] Studying our formulation of single PD solution one can note that the amino acids used are different, one to another in respect of quantitative and qualitative structure. In fact, it contains at relatively high concentration ten indispensable amino acids, histidine, leucine, lysine, HCl, methionine, phenylalanine, threonine, tryptophan, valine, arginine, and glutamine. The total amount of amino acids in our solution is approximately 13.322 g/L and its average mw is about 154.42 with a constant pH 7.25 to 7.45 during its storage and treatment.

[0017] We consider useful to refer here briefly on the role of the numerous dispensable amino acids not included in our PD solution and of the ten indispensable amino acids, selected to be included in the solution indispensable amino acids.

[0018] Glycine, serine, proline, tyrosine, citrulline, ornithine, alanine, aspartic acid, glutamic acid and others dispensable amino acids, were rejected from our single bicarbonate PD solution. The nutritional property of these amino acids is rather poor while they don't have proved specific beneficial properties, except for an interesting cytoprotective action of glycine (6) and the antioxidant property of aspartic and glutamic acids (12). Moreover, serine is not safe, causing rats to die with anorexia, albuminuria, circulatory failure, congestion of liver and lungs (21). It is also doubtful if most of the above amino acids could form glycogen in excess and have an antiketogenic effect. In fact, only alanine, a dispensable amino acid forms glycogen and is antiketogenic, but today glutamine an indispensable amino acid, is more potent and effective than alanine. It considerably enhances carbohydrate metabolism in all human organs with many other beneficial properties (see below).

[0019] The ten amino acids included in our single bicarbonate PD solution are all indispensable which means that they cannot be synthesized in the human organism. Consequently, they may play a potent nutritional role by maintaining energy balance and thus combat and control the malnutrition of chronically uremic patients.

[0020] Histidine was disputed for many years to be an indispensable amino acid. Today, according to many studies, histidine is an indispensable amino acid (22,23). Kopple and Swendsel (24,25) established that histidine might be indispensable in chronically uremic patients, while others confirmed its indispensability (26,27). Newer metabolic studies suggest that histidine at high concentration, is associated with anorexia in animals, which could lead to protein-energy malnutrition. Histidine, the biosynthetic precursor of the neurotransmitter histamine was found in these animals to be elevated in plasma and brain five times above normal. In children, protein-energy malnutrition consistently produces symptoms of depressed food intake, edema, growth failure and psychomotor changes (28), and it must be avoided. On the contrary, in adults at a concentration up to 6 mM/L (as in our solution), histidine enhances nutrition, controls lipid metabolism (26) and maintains hemoglobin levels (29).

[0021] Leucine, lysine, methionine, threonine and valine are all well known indispensable amino acids. They are also the basic potent nutrients various studies showed that the concentrations of these amino acids could be increased manyfold without apparent toxic effects (30-34).

[0022] This is also the case with phenylalanine, and tryptophan. Phenylalanine and tryptophan are indispensable amino acids. Tryptophan is distinguished from other amino acids by its indole ring and it is the first of the amino acids to have been proved indispensable. Both are potent nutrients. They must be used at

low concentrations, up to 2-3 mM/L, as they may cause transient brain disturbances in higher concentrations (35-37).

[0023] The remaining two amino acids, arginine and glutamine, merit more attention, because their roles were extended recently in many fields. Firstly, both amino acids are now classified definitely as indispensable and from this point of view they should have potent nutritional effects (38,39).

[0024] Arginine is the precursor of nitric oxide (NO) (38,39). Its supplementation in the hypercholesterolemic rabbits improves endothelium-dependent vasorelaxation, and shows that this improvement is associated with a reduction in atherogenesis (40-45). Also, in hypercholesterolemic patients arginine corrects the endothelial dysfunction in the vessels of microcirculation (46-49). Of great importance is also the effect of arginine in renal ischaemia. Schamm et al (50) demonstrated that arginine infusion was beneficial after 40 min clamping of the renal arteries, suggesting a regulatory role for the NO pathway in ischemic acute renal failure (ARF). Others shown important protective effect in the renal response to radiocontrast material (51). Also, arginine administration in hypercholesterolemic rats, subjected to high radiocontrast nephrotoxicity, showed important and rapid attenuation (52). However, Tom et al (39) (and others mentioned in this reference) reported recently that NO in excess, generated by irrational administration of arginine, may be directly toxic on cells in ARF and/or by its interaction with superoxide, leading to the oxidant peroxynitrite. The authors substantiated the presence of NO synthetase (NOS) activity in the renal proximal tubules. As already mentioned arginine is the exclusive precursor of NO, a substance with a half-life of only a few seconds, and of a biological activity of 1-2 min. This activity ranges from disposal of protein metabolic waste, muscle metabolism, vascular tone regulation, immune system function, RNA synthesis, and on the release of numerous hormones (catecholamines, glucocorticoids, glucagon, growth hormone, insulin, prolactin, somatostatins), using different pathways, the disturbance of which may produce detrimental effects. Dietary intervention with arginine may improve certain pathological states, such as diabetic nephropathy, cyclosporin A toxicity, salt-sensitive hypertension, ureteral obstruction, kidney hypertrophy due to high-protein feeding, and glomerular thrombosis due to administration of lipopolysaccharides (38,39,53). In order to reduce or even eliminate the detrimental effects and to take advantage of its beneficial effect, we considered necessary, to lower arginine concentration around 3 mM/L in the single bicarbonate PD solution.

[0025] Glutamine is the most abundant free amino acid in plasma and in intracellular pools in all tissues. In humans, glutamine accounts for approximately 20% of all amino acids. Glutamine for many years was neglected but now merits our attention, because during the last decade it was the topic of authentic skilful researches by using isotopic techniques in human providing important and useful information. Today glutamine to be reported as the most important indispensable amino acid (54-57). A brief relation of their properties is needed. Glutamine is the precursor of proteins and nucleotides; it is the substrate and the stimulator of glucogen and gluconeogenesis in all organs and tissues, while alanine gluconeogenesis is poor and takes place in liver only; it is also the regulator of carboxydrate metabolism and antiketotic; the stimulator for the formation of lipids; glutamine suppresses the proteolysis in liver and in skeletal muscles, stimulates their proteins synthesis and it is a strong nutrient (58-71). Finally, glutamine, as also aspartic acid and glutamic acid, according our studies, is a potent scavenger of ROS (see page 13).

[0026] All components of amino acid-based bicarbonate PD solution are absolutely safe at the indicated concentrations. Disodium hydrogen citrate concentration is trivial and after its entry in circulation, citrate is rapidly metabolized to bicarbonate (72), while, the released calcium and magnesium regain their pure forms. The consumption of disodium hydrogen citrate is trivial (0.708 g) for a 6-hr exchange without any alteration of blood coagulation.

Single bicarbonate HD solution

[0027] The exact formulation of single bicarbonate solution is listed in Table 2 (page 15).

[0028] Beside of the presence into the single bicarbonate HD solution (dialysate) of disodium hydrogen citrate at very low concentration (1.5 mM/L), it contains the required electrolytes and the two dispensable amino acids aspartic acid and glutamic acid at a very low concentration each (0.7 mM/L). These along with bicarbonate render the HD dialysate a potent buffer with a constant pH (7.25-7.45). Moreover, these amino acids, according our previous studies, are potent scavengers (see page 13), endow the HD dialysate with strong antioxidative capability, which may combat effectively ROS. As stated above the concentration of disodium hydrogen citrate as well as that of aspartic acid and glutamic acid are unimportant. Nevertheless, one could argue that, because the volume of single bicarbonate HD dialysate during a HD session is around 100 L, the total amount of disodium hydrogen citrate 35.41 g, aspartic acid 9.33 g and glutamic acid 10.3 g that comes in contact with the artificial membrane is too high. Therefore the question arises if these chemical

components could be shown to be dangerous for patient. Taking into account that the HD dialysate exiting from the dialyser retains over 50% of the initial content i.e. half of the above value and that HD is used three times per week, the entry of disodium hydrogen citrate, aspartic acid and glutamic acid could not exceed 7, 2.5 and 2.75 g respectively each day. Citrate even in large amounts is rapidly metabolized to bicarbonate (see above) and the amount of both amino acids is smaller than that of daily intake with food (12).

Id=Table 2. Columns=3

Title: A single and stable improved bicarbonate solution for hemodialysis

Head Col 1 to 2: Components

Head Col 3: g / 100 L

Sodium chloride NaCl 813.62

Sodium bicarbonate NaHCO₃ 268.80

Potassium chloride KCl 11.18

di-Sodium citrate C₆H₈Na₂O₇·11/2 H₂O 35.41

Calcium chloride CaCl₂·2H₂O 25.70

Magnesium chloride MgCl₂·6H₂O 10.18

L-Aspartic acid C₄H₇NO₄ 9.33

L-Glutamic acid C₅H₉NO₄ 10.30

984.50

Na 140 + Cl 111 + HCO₃ 332 +

K 1.5 + Clt 1.5 + Ca 1.75 +

Mg 0.5 + ASP 0.7 + GLU 0.7 mM/L: 289.65

pH : 7.25 - 7.45 Osmolarity: 287 mOsm

Preparation, sterilisation and stability

[0029] Preparation of both PD and HD solutions was carried out by dissolving chemicals in freshly produced ultrapure water taken by a procedure combining carbon perfusion, reverse osmosis and membrane filtration for retaining bacteria. For preparation of HD solution a hundredfold dry mixture of all needed chemicals (984.5 g, see Table 2) was packed and stored in an air tight plastic bottle at room temperature. Thus may serve for preparing adequate dialysate for a routine HD session, by dissolving its content at 100 liters of freshly ultrapure water made by hand or mechanically.

[0030] PD solution could be sterilized by filtration, and also it can be sterilized by autoclaving, according to the classical method used for sterilization of pure bicarbonate solution i.e. at 100 DEG C for one hour, included in air tight plastic bag.

[0031] Sterilized PD solution protected from light and stored at temperature varying from 10 to 35 DEG C, remained stable for at least six months. No precipitate was observed. All components contained into the single bicarbonate amino acid-based solution remained practically unchanged as the difference between monthly measurements did not exceed +/-1.3%. The pH of the solution, ranged within its initial values (7.25 - 7.45). On the other hand, the non sterilized solution of HD, even exposed on air preserves its initial pH (7.25-7.45) for more than 6 hr. Apparent formation of insoluble neutral calcium and magnesium carbonate were not observed. Bicarbonate dialysate prepared on-line is rapidly deteriorated during the HD session, and the insoluble calcium and magnesium carbonate deposition reappear on the wall of flow path tubes.

Comparative study of single amino acid- bicarbonate solution vs lactate solution with 2.5% glucose in rabbits for assessing ultrafiltration

[0032] Two groups of six rabbits (2.5-3.0 Kg) each received i.p. 40 mL/ Kg of either single bicarbonate amino acid-based solution (osmolarity 369 mOsm) and a currently used lactate solution, containing 25 g/L of glucose (osmolarity 408 mOsm), respectively. After a 6-hr dwell time, the animals were sacrificed and the peritoneal fluid was collected, according to a technique reported elsewhere (6). The fluid was measured and net ultrafiltration was calculated as the difference between the infused and drained volume. The mean value of net ultrafiltration was approximately 25% greater in rabbits received single bicarbonate amino acid-based solution than that with lactate solution (59 +/- 2.66 vs 48 +/- 2.5, p< 0.05), although the osmolarity of the first solution was only 369 mOsm and the average mw of amino acids 154.2 against the second solution with an osmolarity 408 mOsm and a mw of glucose 180. Apparently this increased ultrafiltration may be attributed to either the constant normal pH of the single bicarbonate amino acid-based solution, the presence of amino acids, the absence of glucose and its antioxidant property of PD single bicarbonate solution. Other factors may contribute to increased ultrafiltration. As amino acids accept and donate protons, that is, when

they lose protons become negatively charged or when they gain protons become positively charged, it is possible in either of situations that absorption of given indispensable amino acids is repelled from the peritoneum. High ultrafiltration led us to expect that our PD solution would effectively treat the patients by 2 or 3 exchanges daily.

Comparative study of single bicarbonate dialysate vs bicarbonate on-line dialysate in a volunteer patient for assessing various parameters

[0033] Single bicarbonate HD dialysate was used for a 3-month period in a volunteer patient undergoing regular chronic HD with bicarbonate dialysate prepared on-line for 3 yrs. Biochemical values were compared with those of the last 3-month period. Patient served as his own control. The patient tolerated the single bicarbonate dialysate well, as he did previously with the bicarbonate dialysate prepared on-line. There were no significant differences in the following parameters: urea, creatinine, hematocrit, hemoglobin, Na, K, P, Cl, Ca, Mg and PaCO₂. An insignificant increase of pH and HCO₃ values before and after HD with single HD dialysate was observed, while a reduction ($p < 0.05$) of the oxidative activity of serum was noted. This could be attributed to the scavenging property of aspartic acid and glutamic acid, which may attenuate the rate of atheromatosis in chronic uremic patient.

References

1. Yatzidis H: A new stable bicarbonate dialysis solution for peritoneal dialysis. Preliminary report. *Perit Dial Int* 1991; 11: 224-227.
2. Yatzidis H: A new single bicarbonate CAPD solution. In LaGreca G, Ronco G, Feriani M, Chiaramonte S, Conz P (Eds) *Peritoneal Dialysis*. Milano. Wichting Editore, 1991 pp 151-157.
3. Yatzidis H: Hemodialysis with a new single stable bicarbonate dialysate. *Nephron* 1993; 64: 27-31.
4. Yatzidis H: Enhanced ultrafiltration in rabbits with bicarbonate glycylglycine peritoneal dialysis solution. *Perit Dial Int* 1993; 13: 302-306.
5. Yatzidis H: Effect on the peritoneal membrane of rabbits of a single bicarbonate solution containing glycylglycine. *Adv Perit Dial* 1994; 10: 251-255.
6. Yatzidis H, Dombros NV, Digenis GE: On the usefulness of glycylglycine in hemodialysis and peritoneal dialysis solutions. *ASAIO J* 1996; 42: 984-992.
7. Slingeneyer A, Pryzbylski C, Rolland R, Mion C: A new bicarbonate solution for CAPD (Abstract). *Perit Dial Int* 1993; 13 (Suppl 1): 57.
8. Slingeneyer A, Faller B, Michel C, Pryzbylski C, Rolland R, Mion C: Increased ultrafiltration capacity using a new bicarbonate CAPD solution (Abstract). *Perit Dial Int* 1993; (Suppl 1): 57.
9. Pedersen FB: Biocompatibility studies with bicarbonate-based solution. *Adv Perit Dial* 1994; 10: 245-250.
10. Fougeray S, Slingeneyer A, Bastide JM, Mion C: Dialysis solutions buffered with lactate or bicarbonate: in vitro comparison of two dialysis solutions on human peritoneal cell growth from ESRD et non-ESRD patients. *Adv Perit Dial* 1994; 10: 235-240.
11. Fischer F-P, Schenk U, Klefer T, Hubel E, Thomas S, Yatzidis H, Mettang T, Kuhlmann U: In vitro effects of bicarbonate versus lactate buffered CAPD fluid on PMO function. *Am J Kidney Dis* 1995; 24: 924-933.
12. Yatzidis H: New single bicarbonate/ L-aspartic acid/ L-glutamic acid/ glucose (Bi/AAGA/GLU) solutions of physiological pH and variable osmotic pressure for peritoneal dialysis (May 18, 1999 Number of patents 1003136, 1003134 and 1003133).
13. Dudrick SJ, Wilmore DW, Vars HM, Rhoads JE: Long-term total parenteral nutrition with growth, development, and positive nitrogen balance. *Surgery* 1968; 64: 134-142.
14. Gjessing J: Addition of amino acids to peritoneal dialysis fluid: *Lancet* 1968; 2: 812.
15. Oreopoulos DG, Crassweller P, Katirtzoglou A, Ogilvie R, Zellerman G, Rodella H, Vas SI: Amino acids as an osmotic agent (instead of glucose) in continuous ambulatory peritoneal dialysis; in Legrain M (Ed) *CAPD*. London, Oxford-Princeton 1980, pp 335-340.
16. Osborne TB, Mandel LB: Amino acids in nutrition and growth. *J Biol Chem* 1914; 17: 325.
17. Rose WC: The amino acid requirement of adult man. *Nutr Abstr Rev* 1957; 27: 631-47.
18. Irwin MI, Hegsted DM: A conspectus of research on amino acid requirement of man. *J Nutr* 1971; 101: 539-566.
19. Laidlaw SA, Kopple JD: Newer concepts on the indispensable amino acids. *Am J Clin Nutr* 1987; 48: 593-605.
20. The Merck Index: Twelfth Edition 1996, MERCK & C.O., INC, Whitehouse Station, NJ, pp 132.
21. Fishman WH, Artom C: Serine injury. *J Biol Chem* 1942; 145: 345-349.
22. Nasset ES, Gatewood, VH: Nitrogen balance and hemoglobin of adult rats fed amino acid diets low in L- and D-histidine. *J Nutr* 1956; 53: 163-176.
23. Stifel FB, Herman RH: Is histidine an essential amino acid in man? *Am J Clin Nutr* 1972; 25: 182-185.

24. Kopple JD, Swendsel ME: Evidence that histidine is an essential amino acid in normal and chronically uremic man. *J Clin Invest* 1975; 55:881-891.
25. Kopple JD, Swendsel ME: Effect of histidine intake on plasma and urine histidine levels, nitrogen balance and N-methyl histidine excretion in normal and chronically uremic men. *J Nutr* 1981; 111: 931-942.
26. Terry BE, Yamanaka WK, Anderson HL, Wixom RL: Total parenteral nutrition with selective histidine depletion in man. II Hematological, lipid and hormonal responses. *Am J Clin Nutr* 1977; 30: 900-909.
27. Cho ES, Anderson HL, Wixom RL, Hanson KC, Krause GF: Long-term effects of low histidine intake on men. *J Nutr* 1984; 114: 369-384.
28. Mercer LP, Dodds SJ, Weller MD, Dunn JD: Histidine histamine, and neuroregulation of food intake: a review and hypothesis. *Nutrition* 1990; 6:273-277.
29. Giordano C, De Santo NG, Rinaldi S, Pascale C, Pluvio M: Histidine and glycine essential amino acids in uremia. In: Kluthe R, Berlyne G, Burton B (eds). *Uremia, an international conference on pathogenesis, diagnosis and therapy*. Stuttgart: George Thieme Verlag, KG 1972; 138-143.
30. Irving MI, Hegsted DM: A conspectus of research on amino acid requirements of man. *J Nutr* 1971; 101: 539-568.
31. Fellows FCI, Lewis MHR: Lysine metabolism in mammals. *Biochem J* 1973; 136:329-334.
32. Goulet O, DePotter S, Salas J, Robert J-J, Rongier M, Hariz MB, Koziet J, Desjeux J-F, Rigour C, Darmaun D: Leucine metabolism at graded amino acid intakes in children receiving parenteral nutrition. *Am J Physiol* 1993; 265: (Endocrinol Metab, 28): E 540-E 546.
33. Ballevre O, Prugnaud J, Houlier ML, Arnal M: Assessment of threonine metabolism in vivo by gas chromatography/mass spectrometry and stable isotope infusion. *Anal Biochem* 1991; 193: 212-219.
34. Kamoun P: Valine is a precursor of propionyl-CoA. *TIBS* 17 may 1992 (Textbook Errors) pp 175-176.
35. Young SN: Use of tryptophan in combination with other antidepressant treatment: A review. *J Psychiatry Neurosci* 1991; 16: 241-269.
36. Batshaw ML, Valle D, Bessman SP: Unsuccessful treatment of phenylketonuria with tyrosine. *J Pediatr* 1981; 99: 159-160.
37. Wurtman RJ: Nutrients that modify brain function. *Sci Am* 1982; 246: 50-59.
38. Reyes AA, Karl IE, Klahr S: Role of arginine in health and in renal diseases. *Am J Physiol* 1994; 267 (Renal Fluid Electrolyte Physiol 36): F 331-346.
39. Torné LA, Yu L, deCastro I, Campos SB, Seguro AC: Beneficial and harmful effects of L-arginine on renal ischaemia. *Nephrol Dial Transplant*, 1999; 14: 1139-1145.
40. Rossitch E, Alexander E, Black PM, Cooke JP: L-arginine normalizes endothelium function in cerebral vessels from hypercholesteremic rabbits. *J Clin Invest* 1991; 87: 1295-1299.
41. Cooke JP, Singer AH, Tsao PS, Zera P, Rowan RA, Billingham E: Antatherogenic effects of L-arginine in the hypercholesteremic rabbit. *J Clin Invest* 1992; 90: 1168-1172.
42. Tsao PS, McEvoy LM, Drexler H, Butcher EC, Cooke JP: Enhanced endothelial adhesiveness is attenuated by L-arginine. *Circulation* 1994; 89:2176-2182.
43. Böger RH, Bode-Böger SM, Mügge A, Kienke S, Brandes R, Dwenger A, Frölich JC: Supplementation of hypercholesterolaemic rabbits with L-arginine reduces the vascular release of superoxide anions and restores NO production. *Atherosclerosis* 1995; 117:273-284.
44. Böger RH, Bode-Böger SM, Kienke S, Stan AC, Nafe R, Frölich JC: Dietary L-arginine decreases myointimal cell proliferation and vascular monocyte accumulation in cholesterol-fed rabbits. *Atherosclerosis* 1998; 136: 67-77.
45. Bode-Böger SM, Böger RH, Kienke S, Böhme M, Phivthong-ngam L, Tascas D, Frölich JC: Chronic dietary supplementation with L-arginine inhibits platelet aggregation and thromboxane A2 synthesis in hypercholesterolaemic rabbits in vivo. *Cardiovascular Research* 1998; 37: 758-764.
46. Drexler H, Zehner AM, Meinzer K, Just H: Correction of endothelial dysfunction in coronary microcirculation of hypercholesterolaemic patients by L-arginine. *Lancet* 1991; 338: 1548-1550.
47. Cooke JP, Dzau J, Creager A: Endothelium dysfunction in hypercholesterolemia is corrected by L-arginine. *Basic Res Cardiol* 1991; 86 (Suppl 2): 173-181.
48. Clarkson P, Adams MR, Powe AJ, Donald AE, McCredie, Robinson J, McCarthy SN, Keech A, Calermajer DS, Deanfield JE: Oral L-arginine improves endothelium-dependent dilation in hypercholesterolemic young adult. *J Clin Invest* 1998; 97: 1989-1994.
49. Moncada S, Palmer RM, Higgs EA: Nitric oxide: physiology, pathophysiology and pharmacology. *Pharmacol Rev* 1991; 43: 109-142.
50. Schramm L, Heidbreder E, Schmitt A: Role of L-arginine-derived NO in ischemic acute renal failure in the rat. *Renal Failure* 1994; 16: 555-569.
51. Agmon Y, Peleg H, Greenfeld, Rosen S, Brezis M: Nitric oxide and prostanooids protect the renal outer medulla from radioccontrast toxicity in the rat. *J Clin Invest* 1994; 94: 1069-1075.
52. Andrade L, Campos SV, Seguro AC: Hypercholesterolemia aggravates radioccontrast nephrotoxicity: Protective role of L-arginine. *Kidney Int* 1998; 53: 1736-1742.
53. Yu L, Gengaro PE, Niederberger M, Burke TJ, Schrier RW: Nitric oxide a mediator in rat tubular hypoxia / reoxygenation injury. *Proc Natl Acad Sci USA* 1994; 91: 1691-1695.
54. Bulus N, Cersomimo E, Chisham F, Abumrad NN: Physiology importance of glutamine. *Metabolism* 1982; 38 (Suppl 1): 1-5.
55. Souba WW, Klimberg VS, Plumley DA, Salloum RM, Flynn TC, Bland KI, Copelana EM: The role of

- glutamine in maintaining a healthy gut and supporting the metabolic response to injury and infection. *J Surg Res* 1990; 98: 383-391.
56. Smith R: Glutamine metabolism and its physiologic importance. *IPEN* 1990; 14: 405-445.
 57. Perriello G, Jorde R, Nurjhan N, Stumvoll M, Dailey G, Jenssen T, Bier DM, Gerich JE: Estimation of the glucose-alanine-lactate-glutamine cycles in postabsorptive man: Role of skeletal muscle. *Am J Physiol* 1995; 269: E 443- E 450.
 58. Stumvoll M, Perriello G, Meyer C, Gerich J: Role of glutamine in human carbohydrate metabolism in Kidney and other tissues. *Kidney Int* 1999; 55: 778-782.
 59. Nurjhan N, Bucci A, Perriello G, Stumvoll M, Dailey S, Bier D, Toft I, Jenssen T, Gerich J: Glutamine: A major gluconeogenic precursor and vehicle for interorgan carbon transport in man. *J Clin Invest* 1995; 95: 272-277.
 60. Hankard RG, Darmaun D, Sager BK, D'Amore D, Parsons WR, Heymond M: Response of glutamine metabolism to exogenous glutamine in humans. *Am J Physiol* 1995; 269: E 663-E 670.
 61. Varnier M, Leese GP, Thompson J, Renkle MJ: Stimulatory effect of glutamine on glycogen accumulation in human skeletal muscle. *Am J Physiol* 1995; 269: E 309- E 315.
 62. Kreider M, Stumvoll M, Meyer C, Overkamp D, Welle S, Gerich J: Steady state and non-steady measurement of plasma glutamine turnover in human. *Am J Physiol* 1997; 272: E 621- E 627.
 63. Perriello G, Nurjhan N, Stumvoll M, Bucci A, Welle S, Dailey G, Bier DM, Toft I, Jenssen TG, Gerich JE: Regulation of gluconeogenesis by glutamine in normal postabsorptive human. *Am J Physiol* 1997; 272: E 437- E 445.
 64. Stumvoll M, Meyer C, Kreider M, Perriello G, Gerich J: Effects of glucagon on renal and hepatic glutamine gluconeogenesis in normal postabsorptive human. *Metabolism* 1998; 47: 1227-1232.
 65. Lavoinne A, Baquet A, Hue L: Stimulation of glycogen synthesis and lipogenesis by glutamine in isolated rat hepatocytes. *Biochem J* 1987; 248: 429-437.
 66. Cercosimo E, Williams P, Hoxworth B, Lacy W, Abumrad N: Glutamine blocks lipolysis and ketogenesis of fasting. *Am J Physiol* 1986; 250: E 248-252.
 67. Wu G, Thompson JR: The effect of glutamine on protein turnover in chick skeletal muscle in vitro. *Biochem J* 1990; 265: 593-598.
 68. Pösö A, Schworer C, Mortimore G: Acceleration of proteolysis in perfused rat liver by deletion of glucogenic amino acids: Regulatory role of glutamine. *Biochem Biophys Res Commun* 1982; 107: 1433-1439.
 69. Seglen PO, Gordon PB, Poli A: Amino acid inhibition of the autophagic/lysosomal pathway of protein degradation in isolated rat hepatocytes. *Biochem Biophys Acta* 1980; 630: 103-118.
 70. Mac Lennan P, Smith K, Weryk B, Watt P, Renkle M: Inhibition of protein breakdown by glutamine in perfused rat skeletal muscle. *FEBS Lett* 1988; 133-136.
 71. Jepson M, Bates P, Broadbent P, Pelf J, Millward D: Relationship between glutamine concentration and protein synthesis in rat skeletal muscle. *Am J Physiol* 1988; 255: E 166- E 172.
 72. Ismail N, Brouillette JR, Mujais SK: Hemodialysis Technology. In: *Clinical Nephrology, Dialysis and Transplantation* (Eds Malluche HH, Sawaya BP, Hakim RM, Sayegh MH 1999 Deutscher Verlag Dr Karl Feistle, München pp 1-37.

Data supplied from the esp@cenet database - 12

Claim

1. The newer proposed two single and stable solutions for peritoneal dialysis (PD) and hemodialysis (HD) contain the needed electrolytes, the bicarbonate and at very small concentration (1.5 mM/L) disodium hydrogen citrate. Besides, the PD solution contains the following ten indispensable amino acids, L-istidine, L-leucine, L-lysine, L-methionine, L-phenylalanine, L-threonine, L-tryptophane, L-valine, L-arginine and L-glutamine, while the HD solution contains the two dispensable amino acids, L-aspartic acid and L-glutamic acid.
2. Both, PD and HD solutions have the same stable pH 7.25-7.45 arising in PD solution from the ten indispensable amino acids and bicarbonate, and in HD solution from the two dispensable amino acids and bicarbonate. These render PD and HD solution authentic buffers. Studies in our laboratory proved that at pH 7.25-7.45, citrate even at the very small concentration of 1.5 mL/L form by its chelating capability with calcium and magnesium stable soluble complexes, excluding so the appearance of neutral insoluble calcium and magnesium carbonate salts.
3. The soluble complexes of PD and HD solutions during their use are transferred in the circulation of patient where citrate rapidly metabolized in liver to bicarbonate, and the released calcium and magnesium, regain their pure forms. Except the role of amino acid in the formation of pH 7.25-7.45 mentioned in claim 2, the ten indispensable amino acids of PD solution improve disturbed nutrition of patients, adjust the removal of water balance via the peritoneum, and as L-glutamine is potent scavenger, neutralizes the harmful reactive oxygen species (ROS). These exist in excess, causing the damage of peritoneum and all cells and tissues. Also, L-aspartic acid and L-glutamic acid, potent scavengers according our studies, neutralize ROS in patient undergoing hemodialysis by using HD solution.
4. PD and HD solutions are prepared by dissolving chemicals in freshly produced ultrapure water by combining, carbon perfusion, reverse osmosis and membrane filtration for retaining bacterials. PD dialysis solution could be sterilized by filtration, but also by autoclaving, using the classical method for sterilization of pure bicarbonate solutions at 100 DEG C for one hour in air tight plastic bag. The sterilized PD solution protected from light and stored at a temperature varying from 10 DEG to 35 DEG C remains stable for at least six months.
5. HD solution preparation is performed by dissolving a hundredfold dry mixture of all needed chemicals (984.5 g) at 100 L of freshly ultrapure water made hand or mechanically, just before HD session.
6. HD solution, even exposed in air preserves its initial pH (7.25-7.45) for more than 8 hr after its preparation. During this time formation of insoluble neutral calcium and magnesium carbonate salts never were observed. On the contrary bicarbonate solution prepared on-line is rapidly deteriorated during the HD session with visible depositions on the inside wall of flow path tubes.
7. PD solution vs standard lactate solution with 25 g/L glucose in rabbits, and HD solution vs on-line prepared HD bicarbonate solution in a human volunteer, compared as above, proved the significant superiority of newer solutions in terms to ultrafiltration, pH, acid-base control and oxidative activity.

Data supplied from the esp@cenet database - 12